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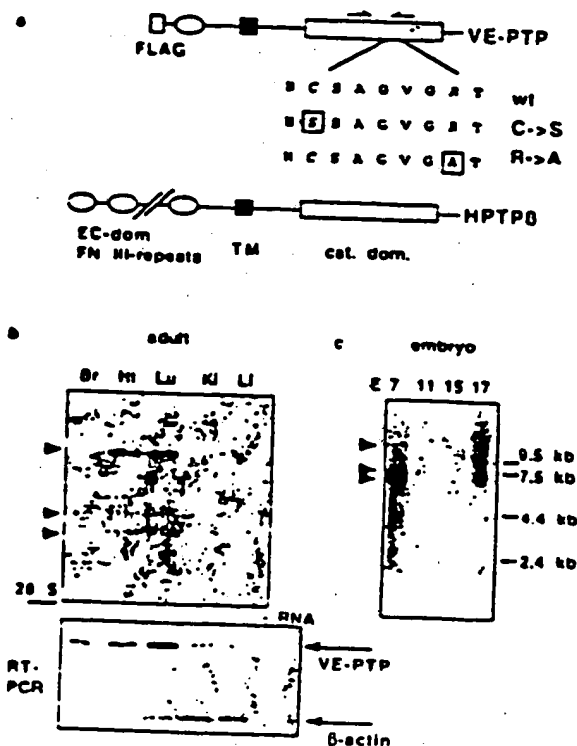
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(54) Title: INTERACTION OF VASCULAR-ENDOTHELIAL PROTEIN-TYROSINE PHOSPHATASE WITH THE ANGIOPOIETIN
RECEPTOR TIE-2

(57) Abstract

Use of vertebrate vascular-endothelial protein tyrosine
phosphatases (i.e. murine phosphatase VE-PTP or human
phosphatase HPTP) or portions thereof for the manufacture
of an agent for monitoring or modulating the activity of the
angiopoietin receptor-type tyrosine kinase Tie-2.



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Interaction of vascular-endothelial protein-tyrosine phosphatase with the Angiopoietin receptor Tie-2

Specification

The present invention relates to a method for monitoring or modulating the activity of the angiopoietin receptor-type tyrosine kinase Tie-2.

10 A key mechanism in the proliferation and differentiation control of all cells are membrane-located receptors, whose activation in many cases is mediated by external factors via phosphorylation of tyrosine residues. The mutation of a series of endothelial cell specific receptor-tyrosine kinases (RTKs) results in lethal phenotypes early during murine embryonal
15 development (Hanahan, Science 277 (1997), 48 - 50; Risau, Nature 386 (1997), 671 - 674). The proliferation and differentiation of endothelial cells depends on two receptor tyrosine kinase systems. The vascular endothelial growth factor (VEGF) is a secreted angiogenic factor and promotes vascularization by activation of its high affinity receptors VEGFR-1 (Flt-1)
20 or VEGFR-2 (Flk-1). The RTKs Tie-1 and Tie-2 are involved in the sprouting and remodelling of the embryonic vascular system. The activity of these kinases is regulated by the recently identified ligands, the angiopoietins.

25 After ligand binding RTKs are activated by phosphorylation on tyrosine residues. Specific protein-tyrosine phosphatases (PTPs) are involved in the fine-tuning of RTK activity. Several classes of PTPs have been identified. However, the biological functions thereof are presently not understood (Neel & Tonks, Curr. Opin. Cell Biol. 9 (1997), 193 - 204; Streuli, Curr. Opin. Cell Biol. 8 (1996), 182 - 188).

30 In a study to identify PTPs in endothelial cells a murine vascular-endothelial protein-tyrosine phosphatase VE-PTP was identified (VE-PTP: a receptor

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protein-tyrosine phosphatase expressed in vascular endothelium, EMBO-FEBS Workshop on Protein Phosphatases and Protein Dephosphorylation, Oxford, UK, September 21 - 26, 1997). Indications for a functional interaction between VE-PTP and a receptor-type kinase have not been
5 described, however. Further, the association of PTPs with their substrates is difficult to determine due to the transient nature of the enzyme substrate association (Flint et al., Proc. Natl. Acad. Sci. U.S.A. 94 (1997), 1680 - 1685).

10 The experiments underlying the present application discovered that VE-PTP is a homolog of the human HPTP β (Krueger et al., EMBO J., 9, (1990), 3241 - 3252), and that it is specifically expressed in endothelial cells both during the embryonal development of mice and in brain capillary vessels of newborn animals. Biochemical analyses using VE-PTP trapping mutants
15 show a specific interaction between the C-terminal part of the molecule which contains the catalytic domain and the RTK Tie-2 but not with the vascular endothelial growth factor receptor VEGFR-2. Moreover, a dephosphorylation of Tie-2 could be detected in the presence of VE-PTP in transiently transfected COS-1 cells. These data identify Tie-2 as a specific
20 substrate for VE-PTP and show that it is a specific modulator of Tie-2 activity.

This result is of high clinical relevance, as Tie-2 holds a key position in angiogenetic processes, the formation of the blood vessel system during
25 embryonal development, the healing of wounds as well as in pathological processes, e.g. tumor development. As VE-PTP shows a specific interaction with Tie-2 and can modulate the tyrosine phosphorylation of the latter, the receptor-protein tyrosine phosphatase is a target both for diagnostic monitoring and for therapeutically influencing the said processes.

30

Thus, a subject matter of the present invention is the use of vertebrate, e.g. mammalian vascular-endothelial protein-tyrosine phosphatases or portions

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thereof for the manufacture of an agent for monitoring or modulating the activity of receptor-type tyrosine kinase Tie-2.

A further subject matter of the present invention is the use of nucleic acids encoding vertebrate, e.g. mammalian vascular-endothelial protein-tyrosine phosphatases or portions thereof for the manufacture of an agent for monitoring or modulating the activity of receptor-type tyrosine kinase Tie-2.

Still a further subject matter of the invention is the use of ligands for vertebrate, e.g. mammalian vascular-endothelial protein-tyrosine phosphatases for the manufacture of an agent for monitoring or modulating the activity of receptor-type tyrosine kinase Tie-2.

The vascular-endothelial protein-tyrosine phosphatases and nucleic acids coding therefor, e.g. genes or cDNA molecules, are obtainable from vertebrate cells, preferably from mammalian endothelial cells, e.g. murine or human cells. Preferably the vascular-endothelial protein-tyrosine phosphatase is selected from murine phosphatase VE-PTP, human phosphatase HPTP β or portions thereof, particularly portions comprising the catalytic domain which is located at the C-terminus of the molecule (Fig. 1a). The nucleic acid sequence and the corresponding amino acid sequence of murine vascular-endothelial protein-tyrosine phosphatase are depicted in SEQ. ID. NO 1 and 2, respectively. The corresponding sequences of the human protein, which were identified by Krueger et al. (supra) are depicted in SEQ. ID. NO 3 and 4.

The polypeptide or a portion thereof is suitable for monitoring or modulating the activity of receptor-type tyrosine kinase Tie-2. In addition to a phosphatase with unmodified sequence of the catalytic domain also mutants thereof, which show a modified, e.g. enhanced binding to Tie-2, e.g. the trapping mutants as depicted in Fig. 2 are suitable for the present

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invention. Particularly mutants, which exhibit an enhanced binding to Tie-2 are well suited for diagnostic and therapeutic applications.

The interaction between the vascular endothelial protein-tyrosine phosphatase and its substrate Tie-2 can also be monitored and/or modulated on the nucleic acid level. To this end nucleic acids, e.g. DNA molecules, RNA molecules or artificial nucleic acid analogs such as peptidic nucleic acids may be used. Preferably these nucleic acids comprise at least 15, particularly at least 20 nucleotides from murine phosphatase VE-PTP gene, human phosphatase HPTP β gene or sequences complementary thereto. These nucleic acids are suitable for the determination of the PTP expression by using known hybridization or/and amplification techniques such as PCR. On the other hand, nucleic acids can be used for the modulation of the VE-PTP expression in the form of antisense constructs or as ribozymes.

A still further aspect of the invention is the use of ligands for vertebrate, e.g. mammalian vascular endothelial-protein tyrosine phosphatases. Examples of such ligands are antibodies, e.g. polyclonal or monoclonal antibodies and antibody fragments. Polyclonal antibodies are available according to known protocols by immunization of test animals with purified VE-PTP, HPTP β or partial fragments thereof, which preferably contain the catalytic domain. From these test animals monoclonal antibodies can be generated in a known manner by using the method applied by Koehler and Milstein. The polyclonal or monoclonal antibodies can also be used in the form of fragments which are obtainable by proteolytic treatment or genetic engineering.

One embodiment of the invention concerns the monitoring or detection of the Tie-2 activity. This detection can be carried out by using known methods, e.g. using labelled polypeptides, nucleic acids or antibodies. A

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further embodiment concerns the modulation of the Tie-2 activity. Thereby a stimulation or a repression of the Tie-2 activity is possible.

Of major importance is the examination or influencing of the interaction
s between VE-PTP and Tie-2 for angiogenesis. Thus the present invention provides means for inducing or for inhibiting vascular growth or remodelling and blood vessel maturation. Particularly, the present invention provides means for inhibiting tumor growth and formation of tumor metastases, e.g. by repressing Tie-2 activity in target cells.

1c

Moreover, the invention is explained by the following figures and sequence protocols.

Fig. 1a shows the schematic representation of VE-PTP, its
15 genetically engineered trapping mutants and HPTP β .

Fig. 1b and c show Northern blot and RT-PCR analyses of VE-PTP
expression in mouse tissues and during mouse embryonic development.

2c

Fig. 2 shows *in vivo* expression analysis of VE-PTP by *in situ*
hybridization.

Fig. 3 shows biochemical interactions of VE-PTP trapping
25 mutants with Tie-2 protein.

Fig. 4 shows selective dephosphorylation of Tie-2, but not
VEGFR-2 by wild-type VE-PTP.

3c Fig. 5 shows a sequence comparison of the C-terminus of
HPTP β with VE-PTP and the translated "mRPTP β "
sequence. Known protein domains are depicted:

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Membrane proximal FN III-domain (blue),
transmembrane domain (red) and catalytic domain
(green). The catalytic center is characterized by a
C(x)₆R-motif.

5 SEQ. ID. NO. 1 and 2 show the nucleotide sequence of VE-PTP cDNA
and the corresponding amino acid sequence.

10 SEQ. ID. NO. 3 and 4 show the nucleotide sequence of HPTP β cDNA
and the corresponding amino acid sequence.

Example 1

15 A PCR screen of a murine brain capillary cDNA library and reverse
transcribed mRNA of bEND5 endothelioma cells to identify endothelial
specific members of the protein-tyrosine phosphatase family was
performed. For PCR, 100 pmol degenerated primers RPTP1 5'-GA(C/T)
TT(C/T) TGG ATG (A/G/T) (G/T) TGG GA-3' and RPTP2 5'-CCI ACI CGI
20 GCI (G/C)(A/T)(A/G) CA(A/G) TGI AC-3' in 50 μ l reactions were used. As
templates 1.25 μ g λ -DNA from mouse P4-10 brain capillary-library
(Schnürch & Risau, Development, 119 (1993), 957 - 968) or 3 μ l of
SuperScript cDNA preparation (GIBCO BRL) from bEND5 mRNA were used.
Isolated 370 bp products were cloned into the vector pCRII (Invitrogen),
25 analysed by restriction cleavage and sequenced on an ABI 370 automated
sequencer (Applied Biosystems).

One of the identified PCR products encodes a polypeptide, designated as
vascular-endothelial protein-tyrosine phosphatase (VE-PTP) which was
identified as murine homolog of the previously described receptor-type
30 protein-tyrosine phosphatase HPTP β (Krueger et al. EMBO J. 9 (1990),
3241 - 3252). VE-PTP and HPTP β belong to the subclass III f receptor-type
PTPs bearing exclusively fibronectin type III-like repeats in the extracellular

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domain and a single catalytic domain in the cytoplasmatic tail (Fig. 1a) (Brady-Kalnay & Tonks, Curr. Opin. Cell. Biol. 7 (1995), 650 - 657).

Fig. 1a shows a schematic representation of VE-PTP, its genetically engineered trapping mutants C->S, R->A and HPTP β . Rectangles indicate mutated amino acids in the catalytic core. The location of the degenerated primers used in the PCR screen are indicated by arrows (EC-dom., extra-cellular domain; FN III fibronectin-type III-like repeat; cat. dom., catalytic domain).

Example 2

A Northern blot and RT-PCR analysis of VE-PTP expression in mouse tissues and during mouse embryonic development were performed. A 751 bp EcoRI-fragment from VE-PTP part 1, obtained by PCR using primers PrPTP β for 5'-GGA AGA GGT ACC TGG TGT CCA TCA AGG-3' and PrPTP β rev 5'-GGC CGG TCC CTA CGA ATG CTG AGC CGG GCA G-3' deduced from a partial clone of murine "RPTP β " (Schepens et al. Mol. Biol. Reports, 16 (1992)), and cloned in the vector pBS KS(+) (Stratagene), was labelled with α^{32} P-dCTP (Amersham Pharmacia Biotech). For Northern blot analysis 20 μ g of total RNA from mouse tissues (Chomczynski & Sacchi, Analyt. Biochem. 162 (1987), 156 - 159) were loaded on a formaldehyde containing agarose gel and blotted. A mouse embryo mRNA Northern blot was obtained from Clontech and hybridization was carried out according to manufacturer's instructions. Autoradiography was performed at -70° C for 17 d. For semiquantitative PCR 50 μ l reactions containing 2 μ l of reverse transcribed cDNA preparations and 20 pmol of primers betaseq2 5'- CCC TCT CCC TTC CTA CCT GG-3' and betarev 5'- GGC CGG TCC CTA CGA ATG CTG AGC CGG GCA GG-3' were used, giving a 416 bp fragment. 30 cycles PCR was optimized to detect 1 fg of VE-PTP plasmid DNA. β -actin RT-PCR was performed as described (Nakajima-Iijima et al, Proc. Natl. Acad. Sci. U.S.A. 82 (1985), 6133 - 6137).

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Northern blot analysis of VE-PTP expression revealed a major transcript of approximately 11 kB and two additional transcripts of 7.5 and 6 kB. In the adult mouse VE-PTP mRNA was strongly expressed in brain as well as in lung and heart. Very weak expression was detectable in kidney and liver (Fig. 1b). These data were confirmed by semi-quantitative RT-PCR performed with RNA from these organs (Fig. 1b). During embryonic development VE-PTP was weakly expressed at embryonic day E11, expression increased at E15 reaching a maximum at E17 (Fig. 1c). Strong expression was detected at E7, which may result from expression in contaminating maternal tissue as expression in the placenta was observed by *in situ* hybridization analysis as well.

Example 3

An *in vivo* expression analysis of VE-PTP by *in situ* hybridization to frozen sections of mouse embryonic tissues was carried out. The results are shown in Fig. 2. Fig. 2a is a darkfield image of an E12.5 embryo section hybridized with a VE-PTP antisense probe. (NC: neural crest, DA: dorsal aorta). Fig. 2b is a darkfield image and Fig. 2c is a brightfield image of a higher magnification of the vessel indicated in a (asteriks). Fig. 2d - h are sagittal sections of E15.5 embryos hybridized with antisense VE-PTP probes. Fig. 2d is a darkfield image and Fig. 2e a brightfield image of the lung. Fig. 2f is a darkfield image of the head region. Fig. 2g is an E15.5 embryo section hybridized with a VEGFR-2 antisense probe. Fig. 2h - k are vessels in brain sections of P10 mice hybridized with antisense VE-PTP probes. As templates for *in vitro* transcription pCRII (Invitrogen) VE-PTP-1 (370 bp fragment of VE-PTP coding for protein sequence corresponding to aa 1786 - 1913 in HPTP β in pCRII) and pBS VE-PTPpart1 were used. Sectioning of mouse embryos and *in situ* hybridization were performed as described (Breier et al, Development, 114 (1992), 521 - 532).

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At the earliest timepoint analysed (E9.5), expression was detectable in the endothelial cell layer lining the dorsal aortae. During the subsequent developmental stages VE-PTP expression was increased throughout the developing vascular system (Fig. 2a). Strong hybridization signals were visible in endothelial cells forming blood vessels, whereas no specific signals were detected in blood cells or smooth muscle cells surrounding the vessels (Fig. 2b, c). At E15.5 specific signals were detectable in all organs with highest expression in the lung (Fig. 2d,e). Comparison to serial sections hybridized with an antisense probe to VEGFR-2 (Flk-1) as an endothelial cell marker, confirmed the vascular endothelial specific expression pattern of VE-PTP (Fig. 2 f,g). In contrast to the uniform expression levels of VEGFR-2 in different types of embryonic endothelial cells, VE-PTP was more strongly expressed in endothelial cells lining larger, smooth muscle cell invested vessels than those of small capillaries and veins. On brain sections of newborn mice, specific expression of VE-PTP was detectable in brain capillaries as well as in larger vessels (Fig. 2h-k). No specific signals were visible in neuronal or glial cells.

Example 4

The biochemical interactions of VE-PTP with the receptor tyrosine kinases Tie-2 and VEGFR-2 were investigated using bacterial GST-fusion proteins. The results are shown in Fig. 3.

Fig. 3a demonstrates the results of GST-fusion pull down experiments. GST and GST x VE-PTP R/A fusion protein were incubated with lysates from bEND5 cells. Precipitates were blotted with an anti-Tie-2 antibody and reblotted with an VEGFR-2 specific antibody. (tot. lys.: total lysates of bEND5 cells). pGEX-VE-PTP contains a 1.1 kB 3' part of EST-clone 552065 (Lennon et al., Genomics 33 (1996), 151 -152) coding for the cytoplasmic domain of VE-PTP cloned in pGEX 3T (Amersham Pharmacia Biotech). GST and GST-fusion proteins were expressed in *E.coli* strain TOP10 essentially

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as described (Frangioni & Neel, Anal. Biochem. 210 (1993), 179 - 187). For pull down experiments 10 cm dishes of confluent endothelial cells were pretreated with pervanadate, lysed and incubated with 10 μ g of GST-fusion protein prebound to glutathion-sepharose as described before (Jallal et al., J. Biol. Chem. 272 (1997), 12158 - 12163).

Fig. 3b shows co-immunoprecipitation of VE-PTP trapping mutants (C->S, R->A) with Tie-2. COS-1 cells were transfected with FLAG-tagged VE-PTP and trapping mutants together with Tie-2. Immunoprecipitation was performed with anti-FLAG antibody M2. Precipitates were blotted with a Tie-2 specific monoclonal antibody.

pCMV-FLAG VE-PTP wt, C->S and R->A contain cDNA sequences coding for a polypeptide stretch corresponding to aa 1418-1977 in HPTP β cloned in pCMV-FLAG-1 (Kodak). Trapping mutations C->S and R->A were introduced by PCR mutagenesis using primer Prbetamutcs 5'-TCC GTA GTG CAC TCG AGT GCT GGT GTG-3' and primer Prbetamutra 5'-GCT GGT GTG GGC GCC ACA GGG ACG TTC-3'. COS-1 cells (Gluzman, Cell 23 (1981), 175 - 182) were transfected using the modified calcium phosphate method (Chen & Okayama, Mol. Cell. Biol. 7 (1987), 2745 - 2752). For transfection 10 μ g of pCMV-FLAG derivatives and 2 μ g of expression plasmids coding for the RTKs were used. As control 0.5 μ g of EGFP expression plasmid (Clontech) were cotransfected. Cells were harvested after 2 d of expression. Transfection efficiency was evaluated under fluorescent light and was usually between 30 - 70%.

In mixing experiments of endothelial cell lysates and trapping mutants of the VE-PTP catalytic domain fused to GST, we detected interaction with the Tie-2 receptor, while GST alone did not precipitate Tie-2. The interaction was independent of pretreatment with pervanadate. In these assays coprecipitation of VEGFR-2 was never detectable (Fig. 3a).

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To test for potential substrate interactions with Tie-2 and VEGFR-2 we coexpressed these RTKs with either a FLAG-tagged version of VE-PTP corresponding to aa 1418-1997 of HPTP β , or the respective trapping mutants (Fig. 1a). Physical association was analysed by co-immunoprecipitation using an anti-FLAG-antibody and subsequent blotting of the precipitates with antibodies specific for the respective RTK. In this assay the Tie-2 receptor co-precipitated with both trapping mutants of VE-PTP (C->S, R->A) (Fig. 3b). The wild type phosphatase failed to precipitate Tie-2 efficiently, even though the receptor was expressed at comparable levels. This reduced association of PTPs *in vitro* with their substrates is due to the transient nature of the enzyme substrate association. Unlike Tie-2, VEGFR-2 could neither be co-immunoprecipitated with VE-PTP nor with one of the trapping mutants, even though VEGFR-2 expression was comparable to that of Tie-2.

Example 5

Finally, the phosphorylation state of RTKs was determined in the presence of VE-PTP. Figure 4 shows dephosphorylation of (a) Tie-2 but not (b) VEGFR-2 by wild-type VE-PTP. RTKs were immunoprecipitated with specific antibodies from cotransfected COS-1 cells. Precipitates were blotted with anti-phosphotyrosine antibodies and after stripping reprobed with RTK-specific antibodies.

Tie-2 and VEGFR-2 expression vectors were published previously (Koblizek et al., Curr. Biol. 8 (1997), 529 - 532; Millauer et al., Cell 72 (1993), 835 - 846). Rat monoclonal antibodies against Tie-2 clones 3g1 and 4g8 (Koblizek et al. (1997) supra) and Flk-1 clone 12 α 1 (Kataoka et al., Devel. Growth Diff. 39 (1997), 729 - 740) were used. Immunoprecipitations were performed with 5 μ g of the monoclonal antibodies and immunoblotting with 2 μ g/ml. Polyclonal anti-Flk-1 serum 1D3 (Sugen) was used in a 1:5000 dilution. Monoclonal anti-Flag antibody M2 (Kodak), polyclonal antiserum

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against Tie-2 (Santa Cruz Biotechnology) and monoclonal mouse antibody against phosphotyrosine PY20 (Transduction Labs) were used according to the manufacturer's instructions. Immunoprecipitations and immunoblotting were performed as described before (Esser et al., J. Cell. Biol. 140 (1998), 947 - 959); Jallat et al., J. Cell. Biol. Chem. 272 (1997), 12158 - 12162).

Immunoprecipitates of VEGFR-2 and Tie-2 co-expressed with either the VE-PTP trapping mutants (C->S, R->A) or wt VE-PTP were blotted with an α -phosphotyrosine-specific antibody and then reprobed with antibody specific for the RTK. Only for Tie-2, changes in the phosphorylation status were observed. In the presence of the trapping mutants (C->S, R->A) the receptor was reproducibly more highly phosphorylated than in the controls. This hyperphosphorylation of Tie-2 in the presence of catalytically impaired trapping mutants suggests that physical interaction leads to protection of the receptor from dephosphorylation. In contrast, hypophosphorylation of the Tie-2 receptor was observed in the presence of wt VE-PTP, when compared to vector control (Fig. 4a). No significant changes were detected in the phosphorylation status of VEGFR-2, irrespective of the presence of VE-PTP or its trapping mutants (Fig. 4b). These findings clearly show that Tie-2 is a specific substrate for the endothelial specific phosphatase VE-PTP.

Claims

1. Use of vertebrate vascular-endothelial protein-tyrosine phosphatases or portions thereof for the manufacture of an agent for monitoring or modulating the activity of receptor-type tyrosine kinase Tie-2.
2. The use of claim 1 wherein said phosphatase is selected from murine phosphatase VE-PTP, human phosphatase HPTP β or portions thereof.
3. The use of claim 1 or 2 wherein said portion comprises the catalytic domain.
4. Use of nucleic acids encoding vertebrate vascular-endothelial protein-tyrosine phosphatases or portions thereof for the manufacture of an agent for monitoring or modulating the activity of receptor-type tyrosine kinase Tie-2.
5. The use of claim 4 wherein said nucleic acid comprises at least 15 nucleotides from murine phosphatase VE-PTP nucleic acid, human phosphatase HPTP β nucleic acid or sequences complementary thereto.
6. The use of ligands for vertebrate vascular-endothelial protein-tyrosine phosphatases for the manufacture of an agent for monitoring or modulating the activity of receptor-type tyrosine kinase Tie-2.
7. The use of claim 7 wherein said ligands are selected from antibodies and antibody fragments.
8. The use of any one of claims 1 - 7 for detecting Tie-2 activity.

9. The use of any one of claims 1 - 7 for stimulating Tie-2 activity.
10. The use of any one of claims 1 - 7 for repressing Tie-2 activity.
- 5 11. The use of any one of the previous claims for monitoring or modulating angiogenesis.
12. The use of any one of the previous claims for inducing vascular growth or remodelling and blood vessel maturation.
- 10 13. The use of any one of the previous claims for inhibiting vascular growth or remodelling and blood vessel maturation.
14. The use of any one of the previous claims for inhibiting tumor growth.
- 15 15. The use of any one of the previous claims for inhibiting formation of tumor metastases.

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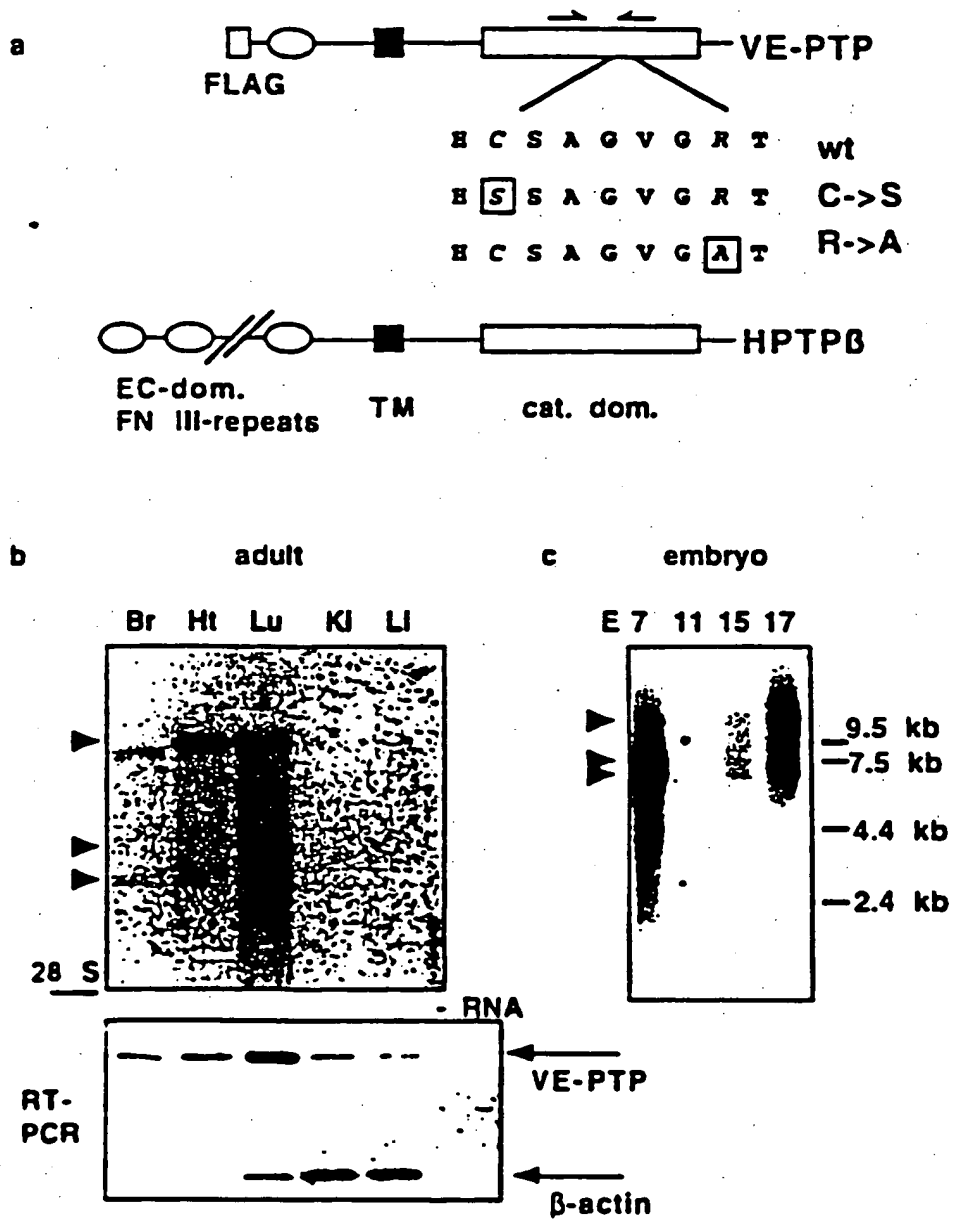


Fig. 1

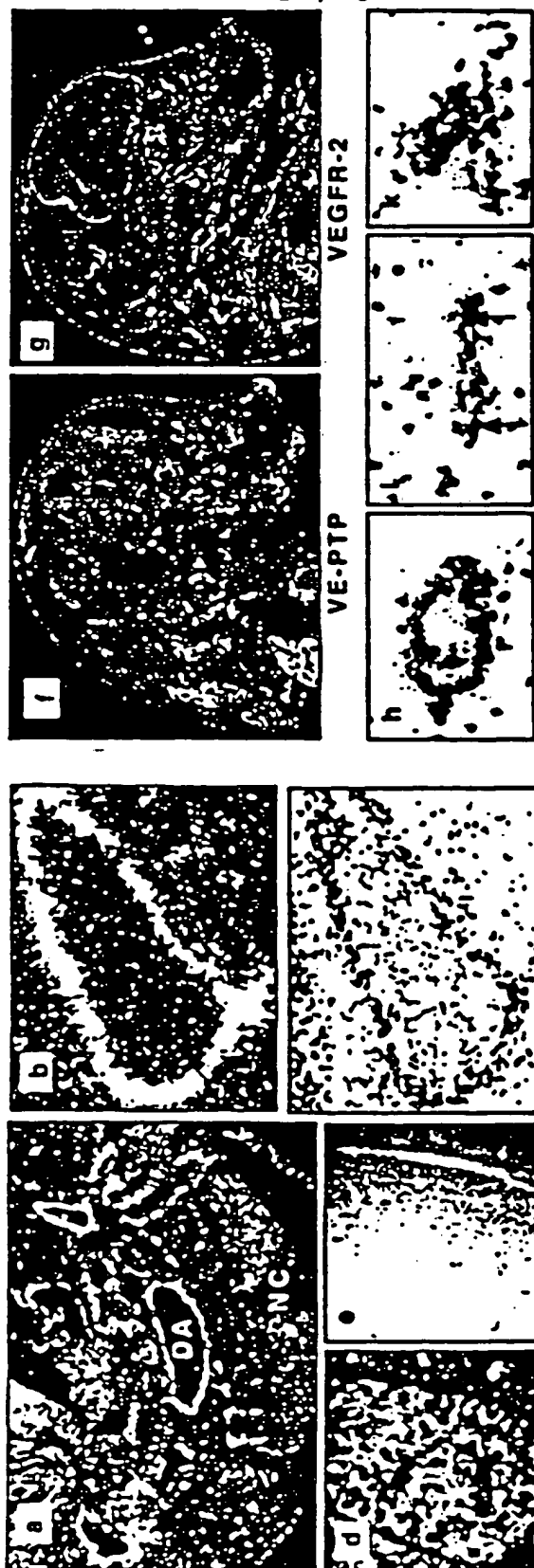


Fig. 2

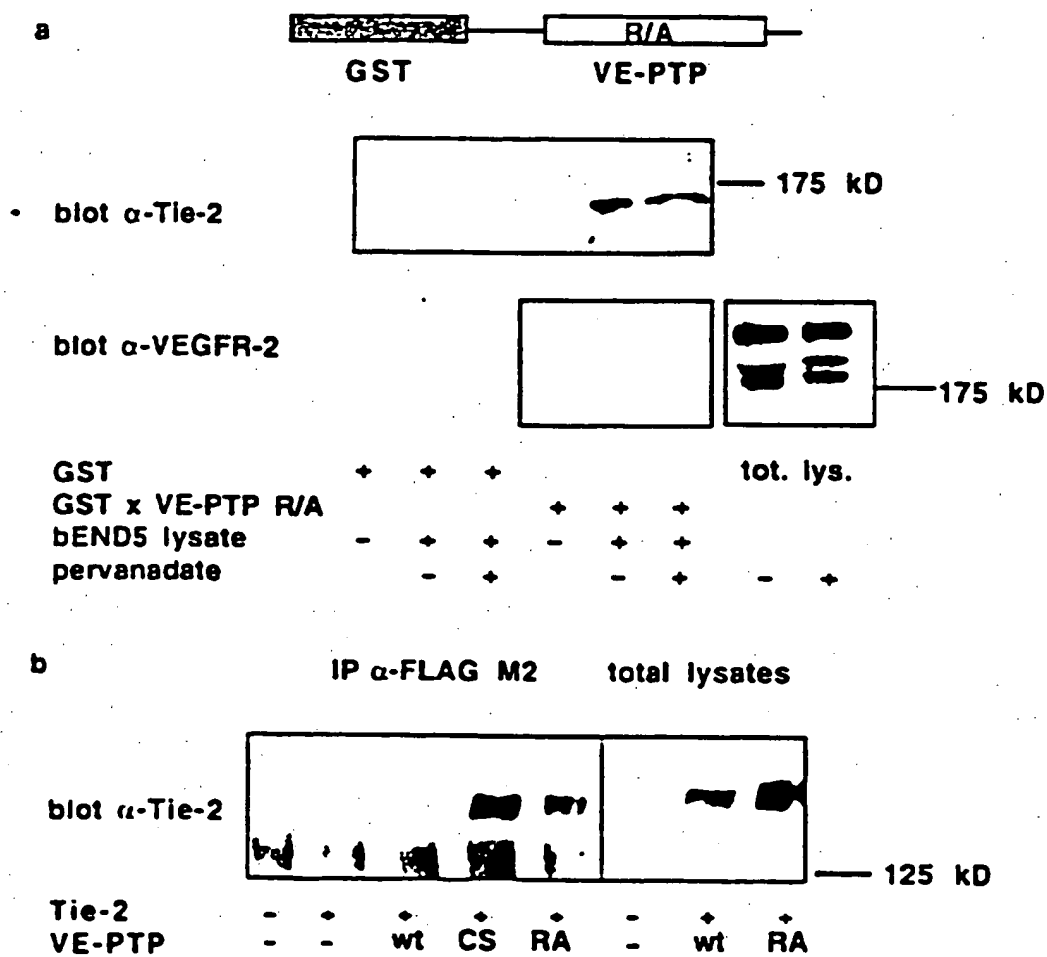


Fig. 3

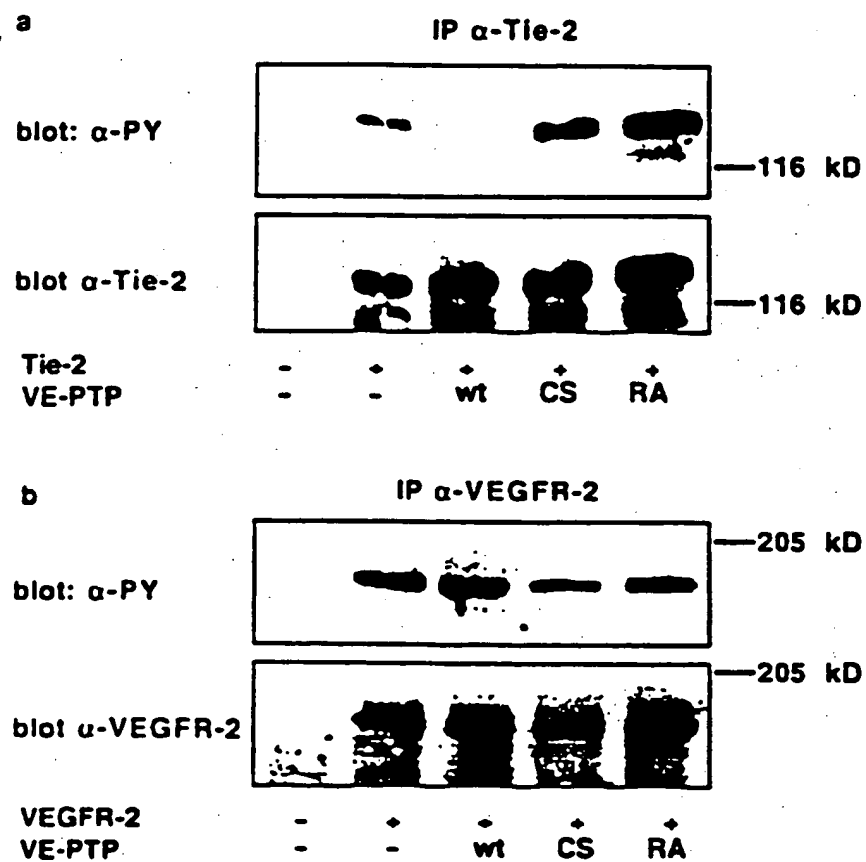


Fig. 4

Fig. 5

HPTPB aal417 . VPHKRYLVS IKVQSAGMTSEVVEDSTIEMDRPPPPPPHIRVNEKDV
 VE-PTP YLVS IKVQSAGMTSEVVEDSTIEMDRPPQPPPHIRVNEKDV
 ,mRPTB SRKRYLVS IKVQSAGMTSEVVEDSTIEMDRPPQPPPHIRVNEKDV

LISKSSINFTVNC SWFSDTNGAVKYFTVVVREADGSDELKPEQQHPLPSYLEYRHNASIRVYQT
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Thr Val Pro Ser Ala Val Lys Asn Ile His Ile Ser Pro Asn Gly Ala			
905	910	915	920
aca gat agc ctg acg gcc aac tgg att cct gcc ggg gga gac gcc gat	2838		
Thr Asp Ser Leu Thr Val Asn Trp Thr Pro Gly Gly Gly Asp Val Asp			
925	930	935	
ccc tac acg gcc tgg gca ttc agt cac agt caa aag gcc gac tct cag	2886		
Ser Tyr Thr Val Ser Ala Phe Arg His Ser Gln Lys Val Asp Ser Gln			
940	945	950	
acc att ccc aag cac gcc ttc gag cac acg ttc cac aga ctg gag gcc	2934		
Thr Ile Pro Lys His Val Phe Glu His Thr Phe His Arg Leu Glu Ala			
955	960	965	
ggg gag cag tac cag att atg att gcc tca gcc agc ggg tcc ctg aag	2982		
Gly Glu Gln Tyr Gln Ile Met Ile Ala Ser Val Ser Gly Ser Leu Lys			
970	975	980	
aat cag ata aat gcc gcc ggg ctg aca gcc cca gca tct gcc caa gga	3030		
Asn Gln Ile Asn Val Val Gly Arg Thr Val Pro Ala Ser Val Gln Gly			
985	990	995	1000
gca att gca gat aat gca tac agc agt tat tcc tta ata gta agt tgg	3078		
Val Ile Ala Asp Asn Ala Tyr Ser Ser Tyr Ser Leu Ile Val Ser Trp			
1005	1010	1015	
caa aaa gcc gcc gcc gcc gca gaa aga tat gat atc ctg ctt cta acc	3126		
Gln Lys Ala Ala Gly Val Ala Glu Arg Tyr Asp Ile Leu Leu Leu Thr			
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gaa aat gga att cct ctg cgc aac aca tca gag cca gcc att att aag	3174		
Glu Asn Gly Ile Leu Leu Arg Asn Thr Ser Glu Pro Ala Thr Thr Lys			
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caa cac aaa tct gaa gat cta aca cca gcc aag aaa tac aag ata cag	3222		
Gln His Lys Phe Glu Asp Leu Thr Pro Gly Lys Lys Tyr Lys Il Gln			
1050	1055	1060	
att cta att gcc agt gga ggc ctc ttc agt aag gaa gcc cag att gaa	3270		

Ile Leu Thr Val Ser Gly Gly Leu Ph	Ser Lys Glu Ala Gln Thr Glu	
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ggc cga aca gtc cca gca gcc gtc acc gac ctg agg atc aca gag aac		3318
Gly Arg Thr Val Pro Ala Ala Val Thr Asp Leu Arg Ile Thr Glu Asn		
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ccc acc agg cac ctg tcc ttc cgc tgg acc gcc tca gag ggg gag ctc		3366
Ser Thr Arg His Leu Ser Phe Arg Trp Thr Ala Ser Glu Gly Glu Leu		
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agc tgg tac aac atc ttc ttg tac aac cca gat ggg aat ctc cag gag		3414
Ser Trp Tyr Asn Ile Phe Leu Tyr Asn Pro Asp Gly Asn Leu Gln Glu		
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Arg Ala Gln Val Asp Pro Leu Val Gln Ser Phe Ser Phe Gln Asn Leu		
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cta caa gcc aga atg tac aag atg gtc att gta acc cac agt ggg gag		3510
Leu Gln Gly Arg Met Tyr Lys Met Val Ile Val Thr His Ser Gly Glu		
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Leu Ser Asn Glu Ser Phe Ile Phe Gly Arg Thr Val Pro Ala Ser Val		
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agc cat ctc agt ggt tcc aat cgt aac acg aca gac agc ctc tgg ttc		3606
Ser His Leu Arg Gly Ser Asn Arg Asn Thr Thr Asp Ser Leu Trp Phe		
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Asn Trp Ser Pro Ala Ser Gly Asp Phe Asp Phe Tyr Glu Leu Ile Leu		
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Tyr Asn Pro Asn Gly Thr Lys Lys Glu Asn Trp Lys Asp Lys Asp Leu		
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agc gag tgg cgt ttc caa gcc ctc gcc ctc gga agc aag tac gtg ctg		3750
Thr Glu Trp Arg Phe Gln Gly Leu Val Pro Gly Arg Lys Tyr Val Leu		
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Trp Val Val Thr His Ser Gly Asp Leu Ser Asn Lys Val Thr Ala Glu		
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Ser Arg Thr Ala Pro Ser Pro Pro Ser Leu Met Ser Phe Ala Asp Ile
 1260 1265 1270

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 Ala Asn Thr Ser Leu Ala Ile Thr Trp Lys Gly Pro Pro Asp Trp Thr
 1275 1280 1285

gac tac aac gac ttc gag cag caa tgg ttg ccc aga gat gca ctc act 3942
 Asp Tyr Asn Asp Phe Glu Leu Gln Trp Leu Pro Arg Asp Ala Leu Thr
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 Val Phe Asn Pro Tyr Asn Asn Arg Lys Ser Glu Gly Arg Ile Val Tyr
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 Gly Leu Arg Pro Gly Arg Ser Tyr Gln Phe Asn Val Lys Thr Val Ser
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 Gly Asp Ser Trp Lys Thr Tyr Ser Lys Pro Ile Phe Gly Ser Val Arg
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acc aag cct gat aag ata caa aac ctg cat tgc cgg cct cag aac tcc 4134
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acg gcc att gcc tgt tct tgg atc cct cct gat tct gac ttc gat ggt 4182
 Thr Ala Ile Ala Cys Ser Trp Ile Pro Pro Asp Ser Asp Phe Asp Gly
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 Tyr Ser Ile Gln Cys Arg Lys Met Asp Thr Gln Glu Val Glu Phe Ser
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ccc cat aag agt tac ctg gtc tcc atc aaa gtc cag tgg gcc ggc atg 4326
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 Thr Ser Glu Val Val Glu Asp Ser Thr Ile Thr Met Ile Asp Arg Pro
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cct cct cca ccc cca cac att cgt gtc aat gaa aag gat gtg cta att 4422

Pro Pro Pr Pr Pro His Ile Arg Val Asn Glu Lys Asp Val Leu Ile	
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Ser Lys Ser Ser Ile Asn Phe Thr Val Asn Cys Ser Trp Phe Ser Asp	
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Thr Asn Gly Ala Val Lys Tyr Phe Thr Val Val Val Arg Glu Ala Asp	
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Gly Ser Asp Glu Leu Lys Pro Glu Gln Gln His Pro Leu Pro Ser Tyr	
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Phe Ala Ser Lys Cys Ala Glu Asn Pro Asn Ser Asn Ser Lys Ser Phe	
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Pro Thr Gln Gln Lys Phe Cys Asp Gly Pro Leu Lys Pro His Thr Ala	
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Tyr Arg Ile Ser Ile Arg Ala Phe Thr Gln Leu Phe Asp Glu Asp Leu	
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Lys Glu Phe Thr Lys Pro Leu Tyr Ser Asp Thr Phe Phe Ser Leu Pro	
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Ile Thr Thr Glu Ser Glu Pro Leu Phe Gly Ala Ile Glu Gly Val Ser	
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Ala Gly Leu Phe Leu Ile Gly Met Leu Val Ala Val Val Ala Leu Leu	
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acc tgc aga caa aaa gtg agc cat ggc cga gaa aga ccc tct gcc cgc	4998

Ile Cys Arg Gln Lys Val Ser His Gly Arg Glu Arg Pro Ser Ala Arg	1645	1650	1655	
ctg agc att cgt agg gat cga cca tta tct gtc cac tta aac ctg ggc				5046
Leu Ser Ile Arg Arg Asp Arg Pro Leu Ser Val His Leu Asn Leu Gly	1660	1665	1670	
cag aaa ggt aac cgg aaa acc tct tgt cca ata aaa ata aat cag tct				5094
Gln Lys Gly Asn Arg Lys Thr Ser Cys Pro Ile Lys Ile Asn Gln Phe	1675	1680	1685	
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Glu Gly His Phe Met Lys Leu Gln Ala Asp Ser Asn Tyr Leu Leu Ser	1690	1695	1700	
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Lys Glu Tyr Glu Glu Leu Lys Asp Val Gly Arg Asn Gln Ser Cys Asp	1705	1710	1725	1720
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Ile Ala Leu Leu Pro Glu Asn Arg Gly Lys Asn Arg Tyr Asn Asn Ile	1725	1730	1735	
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Leu Pro Tyr Asp Ala Thr Arg Val Lys Leu Ser Asn Val Asp Asp Asp	1740	1745	1750	
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Phe Cys Ser Asp Tyr Ile Asn Ala Ser Tyr Ile Pro Gly Asn Asn Phe	1755	1760	1765	
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Arg Arg Glu Tyr Ile Val Thr Gln Gly Pro Leu Pro Gly Thr Lys Asp	1770	1775	1780	
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Asp Phe Trp Lys Met Val Trp Glu Gln Asn Val His Asn Ile Val Met	1785	1790	1795	1800
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Val Thr Gln Cys Val Glu Lys Gly Arg Val Lys Cys Asp His Tyr Trp	1805	1810	1815	
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Pro Ala Asp Gln Asp Ser L L Tyr Tyr Gly Asp Leu Ile Leu Gln Met	1820	1825	1830	
ctt tca gag tcc gtc ctg cct gag tgg acc atc cgg gag tct aag ata				5574

Leu Ser Glu Ser Val Leu Pro Glu Trp Thr Ile Arg Glu Phe Lys Ile			
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Cys Gly Glu Glu Gln Leu Asp Ala His Arg Leu Ile Arg His Phe His			
1850	1855	1860	
tac acg gcg tgg cca gac cat gga gcc cca gaa acc acc cag tct ctg	5670		
Tyr Thr Val Trp Pro Asp His Gly Val Pro Glu Thr Thr Gln Ser Leu			
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acc cag ttc gcg aga acc gcc agg gac tac atc aac aga agc ccg ggc	5718		
Ile Gln Phe Val Arg Thr Val Arg Asp Tyr Ile Asn Arg Ser Pro Gly			
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gcc ggg ccc acc gcg gcg cac tgc agt gcc ggc gcg ggc agg acc gga	5766		
Ala Gly Pro Thr Val Val His Cys Ser Ala Gly Val Gly Arg Thr Gly			
1900	1905	1910	
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Thr Phe Ile Ala Leu Asp Arg Ile Leu Gln Gln Leu Asp Ser Lys Asp			
1915	1920	1925	
ccc gcg gac att tat gga gca gcg cac gac cca aga ctc cac agg gcc	5862		
Ser Val Asp Ile Tyr Gly Ala Val His Asp Leu Arg Leu His Arg Val			
1930	1935	1940	
cac atg gcc cag acc gag tgc cag tat gcc tac cta cat cag tgc gta	5910		
His Met Val Gln Thr Glu Cys Gln Tyr Val Tyr Leu His Gln Cys Val			
1945	1950	1955	1960
aga gat gcc ccc aga gca aga aag cta cgg agt gaa caa gaa aac ccc	5958		
Arg Asp Val Leu Arg Ala Arg Lys Leu Arg Ser Glu Gln Glu Asn Pro			
1965	1970	1975	
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Leu Phe Pro Ile Tyr Glu Asn Val Asn Pro Glu Tyr His Arg Asp Pro			
1980	1985	1990	
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<212> PRT

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Glu Ser Lys Ala Ser Ser His Ser Val Ser Ile Gln Trp Arg Ile Leu
 35 40 45

Gly Ser Pro Cys Asn Phe Ser Leu Ile Tyr Ser Ser Asp Thr Leu Gly
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Ala Ala Leu Cys Pro Thr Phe Arg Ile Asp Asn Thr Thr Tyr Gly Cys
 65 70 75 80

Asn Leu Gln Asp Leu Gln Ala Gly Thr Ile Tyr Asn Phe Lys Ile Ile
 85 90 95

Ser Leu Asp Glu Glu Arg Thr Val Val Leu Gln Thr Asp Pro Leu Pro
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Pro Ala Arg Phe Gly Val Ser Lys Glu Lys Thr Thr Ser Thr Gly Leu
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His Val Trp Trp Thr Pro Ser Ser Gly Lys Val Thr Ser Tyr Glu Val
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Gln Leu Phe Asp Glu Asn Asn Gln Lys Ile Gln Gly Val Gln Ile Gln
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Glu Ser Thr Ser Trp Asn Glu Tyr Thr Phe Phe Asn Leu Thr Ala Gly
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Ser Lys Tyr Asn Ile Ala Ile Thr Ala Val Ser Gly Gly Lys Arg Ser
 180 185 190

Phe Ser Val Tyr Thr Asn Gly Ser Thr Val Pro Ser Pro Val Lys Asp
 195 200 205

Ile Gly Ile Ser Thr Lys Ala Asn Ser Leu Leu Ile Ser Trp Ser His
 210 215 220

Gly Ser Gly Asn Val Glu Arg Tyr Arg Leu Met Leu Met Asp Lys Gly
 225 230 235 240

Ile Leu Val His Gly Gly Val Val Asp Lys His Ala Thr Ser Tyr Ala
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 Phe His Gly Leu Ser Pro Gly Tyr Leu Tyr Asn Leu Thr Val Met Thr
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 Glu-Ala Ala Gly Leu Gln Asn Tyr Arg Trp Lys Leu Val Arg Thr Ala
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 Pro Met Glu Val Ser Asn Leu Lys Val Thr Asn Asp Gly Ser Leu Thr
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 Ser Leu Lys Val Lys Trp Gln Arg Pro Pro Gly Asn Val Asp Ser Tyr
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 Asn Ile Thr Leu Ser His Lys Gly Thr Ile Lys Glu Ser Arg Val Leu
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 Ala Pro Trp Ile Thr Glu Thr His Phe Lys Glu Leu Val Pro Gly Arg
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Tyr Ile Ile Ser Leu Ala Asp Arg Asp Leu Leu Leu Ile His Lys Ser
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Gln Ala Gln Gly Asp Val Glu Phe Tyr Gln Val Leu Leu Ile His Glu
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Ser Phe His Ser Leu Lys Ser Gly Ser Leu Tyr Ser Val Val Val Thr
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Gly Arg Leu Tyr Thr Val Thr Ile Thr Thr Arg Ser Gly Lys Tyr Glu
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Val Ser Val Ser Asn Ser Ala Arg Ser Asp Tyr Leu Arg Val Ser Trp
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 His Thr Phe His Arg Leu Glu Ala Gly Glu Gln Tyr Gln Ile Met Ile
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 995 1000 1005

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Tyr Gly Asp Leu Ile Leu Gln Met Leu Ser Glu Ser Val Leu Pro Glu
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Trp Thr Ile Arg Glu Phe Lys Ile Cys Gly Glu Glu Gln Leu Asp Ala
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His Arg Leu Ile Arg His Phe His Tyr Thr Val Trp Pro Asp His Gly
 1860 1865 1870

Val Pro Glu Thr Thr Gln Ser Leu Ile Gln Phe Val Arg Thr Val Arg
 1875 1880 1885

Asp Tyr Ile Asn Arg Ser Pro Gly Ala Gly Pro Thr Val Val His Cys
 1890 1895 1900

Ser Ala Gly Val Gly Arg Thr Gly Thr Phe Ile Ala Leu Asp Arg Ile
 1905 1910 1915 1920

Leu Gln Gln Leu Asp Ser Lys Asp Ser Val Asp Ile Tyr Gly Ala Val
 1925 1930 1935

His Asp Leu Arg Leu His Arg Val His Met Val Gln Thr Glu Cys Gln
 1940 1945 1950

Tyr Val Tyr Leu His Gln Cys Val Arg Asp Val Leu Arg Ala Arg Lys
 1955 1960 1965

Leu Arg Ser Glu Gln Glu Asn Pro Leu Phe Pro Ile Tyr Glu Asn Val
 1970 1975 1980

Asn Pro Glu Tyr His Arg Asp Pro Val Tyr Ser Arg His
 1985 1990 1995

INTERNATIONAL SEARCH REPORT

Enter the Application No.
PCT/EP 00/03613

A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 C1201/42 A61K38/46 A61K48/00

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C120 A61K

Documentation searched other than minimum documentation is the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

MEDLINE, EPO-Internal, CHEM ABS Data, BIOSIS, WPI Data, PAJ

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevance to claim No.
A	HUANG L ET AL: "GRB2 and SH-PTP2: potentially important endothelial signaling molecules downstream of the TEK/TIE2 receptor tyrosine kinase." ONCOGENE. (1995 NOV 16) 11 (10) 2097-103.. XP002117444 the whole document	1.4,6
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☒ Other documents are cited in the description of this C

☒ Patent family members are cited in annex.

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"Z'" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"Z*" document member of the same patent family

Date of the actual completion of the international search

8 September 2000

Date of making of the international search report

19/09/2000

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INTERNATIONAL SEARCH REPORT

Patent Application No.
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